

# More than movement: the proprioceptive system as a new regulator of musculoskeletal biology

Bavat Bornstein<sup>1</sup>, Nitzan Konstantin<sup>1</sup>, Cristiano Alessandro<sup>2,3</sup>,  
Matthew C Tresch<sup>4,5,6</sup> and Elazar Zelzer<sup>1</sup>



The proprioceptive system is essential for the control of coordinated movement and posture. Thus, traditionally, the study of proprioception has focused on its role in motor control. In this review, we present more recent findings on other, non-traditional functions of this system. We focus on its involvement in musculoskeletal development, function and pathology, including the regulation of spinal alignment, bone fracture repair and joint morphogenesis. We present the hypothesis that the proprioceptive system plays a central role in musculoskeletal biology, and that understanding the underlying molecular mechanisms will promote both basic science and medical innovations. As an example, we discuss recent evidence indicating that Piezo2, a key mechanosensitive ion channel of proprioception, regulates spine alignment and joint development. The presented findings show that the proprioceptive system regulates a wide range of developmental and physiological processes and that its dysfunction may contribute to the etiology of various musculoskeletal pathologies.

## Addresses

<sup>1</sup> Department of Molecular Genetics, Weizmann Institute of Science, Rehovot 76100, Israel

<sup>2</sup> Department of Brain and Behavioral Sciences, University of Pavia, 27100 Pavia PV, Italy

<sup>3</sup> Department of Physiology, Northwestern University, Chicago, IL 60611, United States

<sup>4</sup> Department of Biomedical Engineering, Northwestern University, Evanston, IL, United States

<sup>5</sup> Department of Physical Medicine and Rehabilitation, Northwestern University, Chicago, IL 60611, United States

<sup>6</sup> Shirley Ryan AbilityLab, Chicago, IL 60611, United States

Corresponding author: Zelzer, Elazar ([eli.zelzer@weizmann.ac.il](mailto:eli.zelzer@weizmann.ac.il))

Current Opinion in Physiology 2021, 21:77–89

This review comes from a themed issue on **Proprioception**

Edited by **Tim Cope** and **Leah Bent**

<https://doi.org/10.1016/j.cophys.2021.01.004>

2468-8673/© 2021 Published by Elsevier Ltd.

## Introduction

Proprioception is the sense of the relative position and movement of one's own body parts, the sense of tension or force and of the effort exerted by acting muscles. As such, proprioception is essential for the control of coordinated movement and posture [1,2]. Proprioceptive information is produced by specialized mechanosensory organs termed proprioceptors. In the musculoskeletal system of humans and other terrestrial vertebrates, the predominant types of proprioceptors are the muscle spindle and the Golgi tendon organ (GTO). These types differ in morphology, location, measured input, effect and other traits [3–5]. However, common to both organs is the ability to sense the biomechanical environment, initiate a neural response in specialized sensory afferent fibers, often termed proprioceptive neurons, and modulate local muscle tension by forming the muscle spindle and GTO reflex arches [1]. While the sensory nerve endings of these neurons are located within skeletal muscles and tendons, their cell bodies are found in the dorsal root ganglion (DRG) and they terminate in the spinal cord, where they communicate with motoneurons and interneurons to form reflex arcs. Additionally, these proprioceptive neurons continuously inform the central nervous system (CNS) about the tension, forces and movement of our muscles [1,6,7].

The vertebrate musculoskeletal system is composed of diverse tissue types, including skeletal tissues (bone, cartilage, and joints), muscles, tendons and ligaments. Its development and function were shown to be regulated by cross-tissue communication, which can be mediated by both molecular and mechanical cues [8,9]. The importance of mechanical signals such as muscle contraction for musculoskeletal development is conserved from fish to humans (reviewed in Ref. [9]), indicating that embryonic movement is necessary for this process. In the following, we refer to this type of regulation as non-autonomous, meaning that signals from one tissue, in this case, muscles, affect another tissue, namely skeleton. However, while the non-autonomous effect of muscle activity on skeletal development is well established, it raises questions regarding the nature of the movement that is necessary. Are there certain types of movement that are essential for proper development, or would any movement do? If indeed specific patterns of movement are needed, then abnormal proprioceptive function may be

the cause for various musculoskeletal syndromes and pathologies.

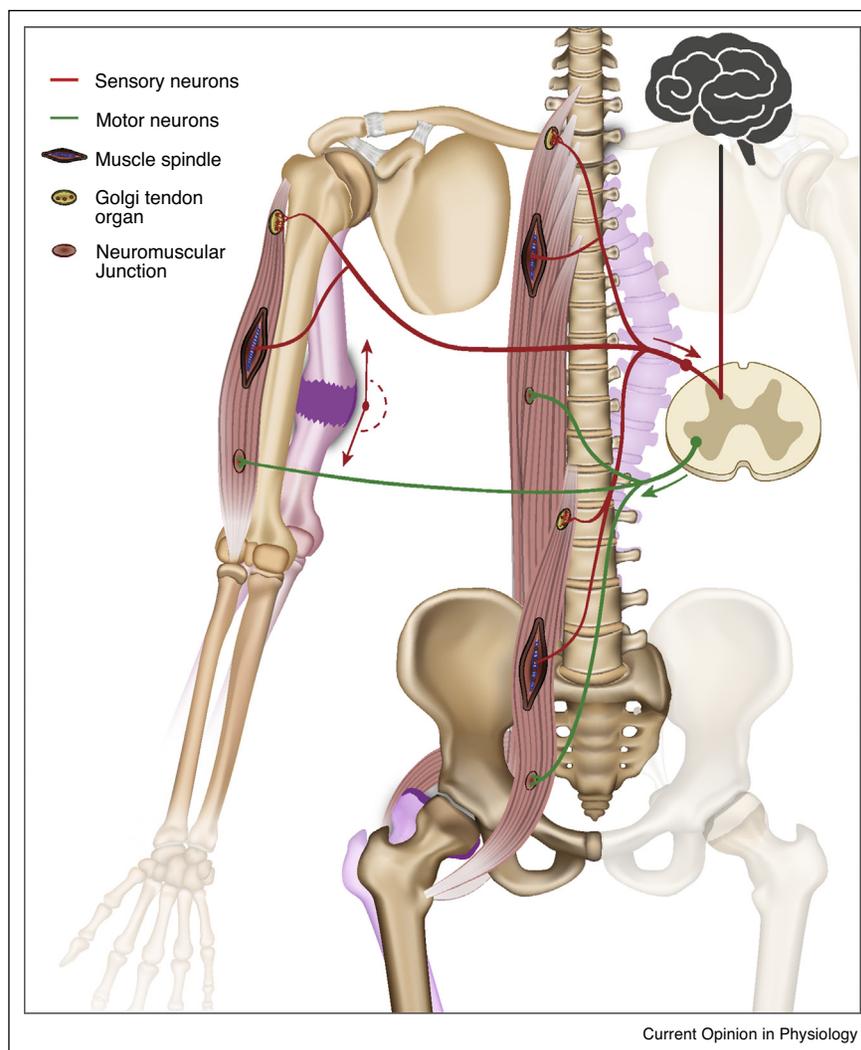
In this review, we present recent findings implicating the proprioceptive system in regulating musculoskeletal biology, as well as in musculoskeletal pathologies. In the first part, we describe the roles of proprioception in spinal alignment, restoration of fractured bones and hip joint morphogenesis (summarized in Figure 1). These findings establish the regulatory role of proprioception in skeletal biology and suggest that mutations in genes associated with proprioception might be involved in musculoskeletal pathologies. This possibility is discussed in the second part of this review, using Piezo2 as a proof of concept. We also present other molecular

component that might function within proprioception to regulate musculoskeletal biology. Finally, we speculate on possible future directions, including studying the involvement of proprioception in other musculoskeletal pathologies.

### Proprioception in musculoskeletal development and function

In this chapter, we provide several examples for the regulation of key aspects in musculoskeletal biology by the proprioceptive system. These examples highlight the possibility that malfunction of the proprioceptive system might be the underlying mechanism in various human musculoskeletal pathologies.

Figure 1



The proprioceptive system regulates skeletal biology.

In the presence of a functional proprioceptive circuitry, properly regulated muscle contractions maintain proper alignment and morphology of skeletal elements. However, loss of proprioceptive circuitry results in skeletal deformity, such as spinal malalignment, aberrant hip joint morphology or impairment in natural reduction of fractured bones (depicted in purple).

### a. Proprioception and scoliosis

The vertebral column is the central axis of the body, serving to support weight and maintain posture while allowing movement. To perform these functions and withstand high stresses created by body weight and by muscle loads, the position and orientation of dozens of vertebrae and intervertebral discs must be tightly maintained. Yet, little is known about the mechanisms that regulate spine alignment. An example for failure of these mechanisms is a condition known as scoliosis, which is defined as a lateral spinal curvature of 10 degrees or more [10]. The most common type of the disease is adolescent idiopathic scoliosis (AIS), which appears during puberty without other skeletal anomalies in around 3% of school-age children worldwide. Although severe cases may interfere with breathing, the mechanisms underlying the pathogenesis of AIS are still largely unknown [11,12].

Since AIS patients have morphologically intact spinal elements, spinal alignment may be regulated non-autonomously. A large body of evidence supports the involvement of the neuromuscular system in the etiology of scoliosis. These include anatomical alterations of motor brain areas, such as the cerebellum, cerebral cortex and vestibular system [13–15]. AIS patients also displayed abnormal somatosensory evoked potentials [16] and asymmetric strength and calmodulin distribution in trunk muscles [17,18]. In addition, neural injuries such as stroke [19] and cerebral palsy [20] are known to be associated with the development of postural imbalance and spine deformity. Finally, in animal models, lesions in dorsal column and posterior horn of the spinal cord [21] or nerve roots [22,23] resulted in scoliosis, demonstrating evolutionary conservation of the association between neuromuscular function and spine alignment.

Interestingly, both stroke [24,25] and cerebral palsy [20] have been shown to affect proprioception, suggesting a mechanistic link between neuromuscular function and the acquired deformity. Indeed, several observations support the notion of impaired proprioceptive function in AIS patients. These include asymmetrical gait [26] and impaired postural and balance control [27,28], as well as abnormal proprioception-related neural responses, such as inability to reproduce joint angle [29], vibratory sensation [30] and size-weight response [31]. Notably, a reduced number of muscle spindles was found in paravertebral muscles of AIS patients [32]. Additionally, the onset of AIS coincides with the maturation of the proprioceptive system, which occurs during the second decade of life [33,34]. While these data indicate a correlation between scoliosis and proprioception, direct evidence for the hypothesized involvement of proprioception and, specifically, of muscle spindles

in the control of spine stability has only recently started to emerge.

The first demonstration of the involvement of proprioception in regulating spine alignment was provided by analyzing *Runx3* knockout (KO) mice, which lack functional proprioceptive neurons [35,36]. These mice developed peripubertal scoliosis without prior vertebral dysplasia or muscle asymmetry [37\*\*]. *Runx3* was also shown to be involved in differentiation of chondrocytes [38,39] and osteoblasts [40,41]. Therefore, a conditional knockout approach was applied to identify the tissue in which *Runx3* function is necessary to prevent scoliosis. Interestingly, while conditional deletion of *Runx3* in skeletal tissues did not affect spine alignment, its deletion from peripheral nervous tissue or specifically from peripheral sensory neurons produced a similar scoliotic phenotype, indicating a non-autonomous role of the nervous system in skeletal alignment. Moreover, deletion of enhancer elements driving *Runx3* expression in proprioceptive neurons induced a similar phenotype, further indicating that proprioceptive neurons are necessary to maintain spinal alignment. A similar phenotype but of reduced severity was seen in *Egr3* KO mice, which lack muscle spindles but not Golgi tendon organs [42,43]. Functional assays revealed a decrease in gait regularity, which was also more pronounced in *Runx3* KO than in *Egr3* KO mice. These findings implicate impaired proprioceptive signaling in acquired scoliosis and suggest that both receptor types are required for this regulatory mechanism.

### b. Proprioception and bone fracture healing

Restoration of bone morphology is essential for successful fracture healing and functional outcome [44,45]. In the orthopedic practice, this procedure is called reduction [46]. However, several pieces of evidence suggest that in nature too, broken bones are rapidly realigned following injury. In humans, humeral birth fractures, even when severely angulated, usually heal well and with little residual deformity without intervention [47]. Studies of primate skeletons revealed high incidence of up to 30% of bone fractures, occurring mostly in early life, which also healed well with minimal residual deformity [48–50].

These findings indicate that during evolution, vertebrates have acquired the ability to actively restore the morphology of fractured bones [51]. Indeed, it was shown that fractured humeri of neonatal mice undergo realignment without any intervention by a process that involves substantial movement of the two fracture fragments, which has been termed natural reduction [52]. This suggests the existences of a mechanism that regulates the restoration of bone morphology and guides realignment [52]. Recent findings showed that natural reduction failed in fractured bones of *Runx3* KO mice, as well as upon conditional deletion of *Runx3* in the peripheral nervous system. Similar to the spine alignment experiments, *Egr3* KO

mice displayed a less severe phenotype. Interestingly, inactivation of muscles surrounding the fracture site also resulted in failed realignment. These findings suggest that both proprioceptor types, as well as muscle contraction, are necessary for the regulation of natural reduction [53\*\*].

Both molecular and mechanical interactions among musculoskeletal tissues have been shown to regulate the development of this system [9,54–57]. For example, mechanical signals generated by skeletal muscles control the formation of the circumferential shape of developing long bones [58] and maintain joint-forming cells committed to their fate [59]. Recently, it was suggested that muscle-derived satellite cells express growth factors that contribute to bone fracture repair [60]. The findings that muscle activity and proprioceptive signaling are involved in the regulation of fracture repair further demonstrates the importance of such cross-tissue interactions.

Bones are mechanosensitive organs [51,61–64] that adjust their morphology [58,65] and composition [66,67] in response to dynamic changes in mechanical loading. Bone and cartilage cells, including, osteoblasts [68] osteocytes [69] and chondrocytes [70,71], have been shown to be mechanosensitive, as is fracture callus [72–74]. The finding of a mechanism that monitors bone integrity and instructs its restoration following trauma through proprioceptive signaling adds a level of nonautonomous mechanosensitivity to the current view of fracture repair.

Interestingly, unlike in most repair processes, we found that natural reduction becomes more effective with age, concurrently with the maturation of the proprioceptive system. In mice, the sensory endings of muscle spindles continue to develop until about postnatal day 40 [75,76], whereas in humans, the performance of proprioception-dependent tasks improves during adolescence [33,34]. These findings support the notion that the observed improvement in natural reduction was due to augmented proprioceptive function.

Thus far, the research of fracture repair has largely overlooked the role of muscle force in bone realignment. Based on our findings, we suggest that the fracture changes the length and tonus of muscles that are attached to the bone, which are detected by muscle proprioceptors. Consequently, proprioceptive signals guide the correction of the position of misaligned fracture fragments rapidly and effectively by inducing asymmetric muscle contraction. Considering the contribution of this mechanism may improve the healing process and its outcome.

### c. Proprioception and joint morphology

#### *Hip dysplasia*

Hip dysplasia, also known as developmental dysplasia of the hip (DDH), is the most common abnormality in human

newborns. It is a spectrum of anatomical abnormalities of the hip joint in which the femoral head is malpositioned relative to its socket, the acetabulum [77]. Ranging from hip instability to dislocation, the condition affects hip biomechanics and, if left untreated, may cause osteoarthritis [78], limping and back pain. One in 100 children is born with hip subluxation or dysplasia and 1 out of 1000 children is born with a dislocated hip; 80% percent of affected children are female [79]. While the causes of hip dysplasia are not fully understood, it is approximately 12 times more likely when there is a family history [<https://hipdysplasia.org/developmental-dysplasia-of-the-hip/causes-of-ddh/>] [80].

The necessity of embryonic movement and muscle-generated forces for joint development has been well-established in various model organisms [37\*\*,77–80], as in the absence of muscle contraction several joints fail to form [78,81,82]. However, this paradigm considers only two situations, namely the existence or absence of movement, disregarding everything in between. This dichotomous view overlooks the intriguing question of whether there are ‘right’ and ‘wrong’ types of movement, that is, if proper joint development requires specific patterns of embryonic movement.

Studies of hip joint morphogenesis in proprioception-deficient mutant mice, which preform uncoordinated movements [37\*\*,43,83], might provide new insight into this question. Adult *Runx3* KO mice, which lack all proprioceptive neurons and, consequently, display severe ataxia, exhibited severe, irregular type hip dysplasia, which was manifested by a prominent cam over the femoral neck [84\*\*]. Additionally, significantly increased acetabular index indicated a shallow acetabulum, whereas significantly reduced central edge angle (CEA) indicated lateralized center of rotation, which reduces joint stability and increases the risk for dislocations. Loss of *Runx3* in the peripheral nervous system, but not in skeletal lineages, led to similar joint abnormalities. Similarly, *Egr3* KO mice, which lack only muscle spindle and display reduced muscle coordination, presented a less severe hip dysplasia phenotype [84\*\*].

The misshapen and incongruent joints observed in mutants for the proprioceptive system regulators *Runx3* and *Egr3* confirm the role of this system in hip joint morphogenesis. Moreover, they support the notion that specific patterns of movement are necessary for proper joint morphogenesis. However, how patterns of movement regulate joint morphogenesis is yet to be determined.

#### *Sensory input controls joint integrity*

The work reviewed in the previous sections, showing that altered neural activity after loss of proprioceptive information results in abnormal joint development, suggests that neural activity plays an important role in the

regulation of joint integrity. Although the significance of maintaining joint integrity is obvious when it fails, as seen in people with ligament ruptures or osteoarthritis [85,86], the role of the nervous system in this regulation is poorly understood [87,88].

The potential role of the CNS in regulating joint integrity has been difficult to evaluate, mainly because of the close relationship between joint mechanics and overall behavior [89]. In any purposeful behavior, the nervous system must act through joints in order to produce limb movement. Thus, any perturbation that impairs neural control of behavior will also alter joint stresses and strains. Conversely, any perturbation affecting neural control of joint integrity might result in damaged joint structures, which would then affect task performance.

A recent set of experiments were able to disentangle these effects and demonstrate that the nervous system directly regulates stresses and strains within joints [90,91,92\*,93\*]. These studies exploited the mechanical properties of the rat knee joint, evaluating whether neural control of quadriceps muscles reflects minimization of mediolateral forces on the patella. Examination of the patterns of correlation amongst quadriceps muscles revealed that the activity of muscles (vastus lateralis, VL; vastus medialis, VM) producing opposing mediolateral forces on the patella were highly correlated with one another, whereas the correlation of these muscles to muscles producing minimal mediolateral patellar forces (vastus intermedius, VI; rectus femoris, RF) was much weaker [91]. The low correlation of VM and VL to VI was surprising, since all three muscles are monoarticular knee extensors and thus have the same contribution to task performance. These results are consistent with a control strategy minimizing the net mediolateral force on the patella.

This possibility was further assessed by a series of experiments that examined how the CNS adapts to perturbations affecting joint integrity. First, the adaptations in quadriceps muscle activations in response to the addition of a lateral load on the patella were measured [93\*]. Consistent with neural control of joint stresses, the ratio between the activation of VL and VM was reduced, thereby reducing net mediolateral patellar forces. In another experiment, adaptations were examined following paralysis of VL [92\*]. If the CNS only regulated task performance, increasing activation of VM would replace the lost knee extension torque and restore limb kinematics. Instead, an increased activation of RF to compensate for paralysis of VL was observed. Although this adaptation strategy complicates restoration of task performance, as RF produces a hip flexion torque that is not produced by VL, it is consistent with regulation of joint stresses, since RF produces minimal mediolateral patellar force.

Taken together, these findings support the hypothesis that the nervous system chooses muscle activation patterns to minimize joint stresses and strains while achieving desired task performance. However, how the nervous system monitors these stresses and strains remains unclear [87,88,94]. Although it is possible that joint afferents provide direct information about the state of internal joint structures [95,96], the experiments on proprioception-deprived mice raise an intriguing possibility that muscle proprioceptors may also play an important role. For instance, the pattern of stretch reflexes amongst quadriceps muscles in the cat is consistent with the minimization of mediolateral patellar forces described in our previous experiments [97]. Deep muscles in the shoulder and hip are rich in proprioceptors, suggesting that they might act as sensory structures that signal the position of the femoral or humeral head relative to their sockets [98,99]. Joint loading might be monitored by GTO activity, with the nervous system using ‘internal models’ of how muscle forces affect joint structures to estimate joint stresses and contact forces.

Although the role of muscle proprioceptors in the production of behavior and task performance has been examined extensively [1,2], there are relatively few studies examining the potential role of muscle proprioceptors in regulating joint integrity [94,100]. Future studies in animals lacking muscle proprioceptors are needed to establish the connection between proprioception and joint stability.

In summary, the above examples provide strong evidence for the involvement of the proprioceptive system in regulation of skeletal development and pathology. Therefore, understanding the molecular components of the proprioception system might provide new insight into the mechanisms underlying some of these pathologies.

### **Molecular components mediating proprioception**

While many of the molecular components regulating different aspects of neurobiology have been extensively studied and identified, the molecular mechanisms that regulate the development and function of proprioceptors have attracted less attention. In order to translate the findings that the proprioceptive system regulates musculoskeletal biology into therapeutic applications, it is necessary to decipher these mechanisms. Indeed, several groups have recently started to provide this information [101–103]. In this chapter, we discuss several molecular components within proprioceptors that might function to regulate musculoskeletal biology.

#### **a. Mechanotransduction ion channels in proprioceptive neurons**

Proprioceptive neurons transduce mechanical strain, experienced by muscles and joints, into electrical signals

[1]. While the molecular mechanisms underlying proprioception transduction are largely unknown, it was shown that the major mechanotransducer of mammalian proprioceptors is the ion channel Piezo2 [83].

*Piezo2: the key mechanosensitive ion channel of proprioception*

Piezo1 and Piezo2 are calcium-permeable mechanosensitive ion channels. They are larger in size than other ion channels and their unique structure includes a series of four transmembrane helical bundles termed Piezo repeats, which together compose flexible propeller blades [104–107]. In recent years, this family of ion channels has been implicated in a variety of developmental and physiological processes. Piezo2 was shown to be expressed in lung [108], where it is important for the sensation of airway dilatation [109], in the gastrointestinal tract [110] and in skin, where this channel is involved in sensation of fine touch [111]. Another tissue where both Piezo1 and Piezo2 were found to be expressed is articular cartilage. It was shown that synergy between the two channels provides articular chondrocytes with mechanotransduction ability under high-strain mechanical stress, which is potentially deleterious to these cells [112].

In addition, Piezo2 was shown to be expressed by dorsal root ganglia (DRG) neurons, including proprioceptive mechanosensing neurons ending in muscle spindles and GTOs [83]. Loss of Piezo2 in proprioceptive neurons of mice resulted in severely uncoordinated body movements and abnormal limb positions, leading the authors to suggest that Piezo2 is the main mechanotransducer of mammalian proprioceptors [83].

In humans, mutations in the *PIEZO2* gene result in a variety of pathologies such as proprioception defects, skeletal abnormalities including scoliosis, hip dysplasia and arthrogryposis, a congenital contracture of multiple joints, neonatal respiratory insufficiency and muscle weakness [113–116]. Considering the wide range of tissues that express this gene, it is difficult to associate between phenotype and affected tissue in humans with *PIEZO2* mutations. In mice, however, this question can be addressed directly by creating tissue-specific knockout of *Piezo2*.

Given the observation of skeletal phenotypes in humans, the autonomous role of *Piezo2* was evaluated in mice by blocking its expression in chondrogenic or osteogenic lineages. However this deletion did not lead to alterations in skeletal morphology [84<sup>\*\*</sup>,117]. By contrast, loss of *Piezo2* from proprioceptive neurons led to spinal malalignment and hip dysplasia [84<sup>\*\*</sup>]. *In vivo* CT scans revealed that by P60, these mice developed scoliosis as well as kyphosis. Histological examination and morphometric analysis showed that the spinal deformities were not caused by aberrant development or morphogenesis of

vertebrae or surrounding tissue. Morphological abnormalities in the hip joint included elevated acetabular index and flattened type dysplasia, loss of joint congruency and a femoral cam deformity. These findings establish the proprioceptive system as a regulator of spine alignment and joint development and identify Piezo2 as a key molecular component in this regulatory mechanism. Moreover, they also reinforce the need to better understand the molecular mechanism of proprioception and focus the attention on PIEZO2 regulation and signaling.

To identify proteins that interact with Piezo2, a mass spectrometry analysis was performed on Piezo2 in murine DRG somatosensory neurons [118]. This analysis revealed several candidates for Piezo2 regulation, including myotubularin-related protein 2 (Mtmr2) [118]. Subsequent work has shown that Mtmr2 regulates Piezo2 signaling by decreasing Piezo2-mediated rapidly adapting mechanically activated (RA-MA) currents [119]. Interestingly, Mtmr2 has been implicated in Charcot-Marie-Tooth type 4B1 disease (CMT4B1), a peripheral neuropathy characterized by neuronal demyelination, muscle weakness and sensory loss [120]. In severe cases, various degrees of skeletal deformities such as scoliosis, pectus carinatum, hand clawing, femoral anteversion, internal tibial torsion, in-toeing, and severe equinovarus were also observed [121].

It was recently shown that the regulatory channel TMEM150C/Tentonin3 can regulate Piezo1/2 and TREK-1 ion channels [122,123]. Moreover, TMEM150C/Tentonin3 was shown to be co-expressed with Piezo2, suggesting that it can regulate Piezo2 activity in proprioceptive neurons. Correspondingly, TMEM150C/Tentonin3 mutant shows impaired motor coordination [123]. It remains to be seen whether this fine regulation of proprioceptive sensing, which still affects motor behavior, also affects skeletal alignment.

*Are there other mechanosensitive ion channels?*

Notwithstanding the central role of Piezo2 in mediating proprioceptive signals, it is more than likely that other mechanosensitive channels also contribute to or modulate the stretch-activated response of proprioceptive neurons. Indeed, proprioceptive sensory neurons express other stretch-sensitive channels, such as epithelial sodium channels (ENaCs) including the acid-sensing ion channels (ASICs) [3,124–126]. Knockout of *Asic3* in mice impairs mechanotransduction of proprioceptive DRG neurons and leads to deficits in motor tasks that rely specifically on proprioception, such as grid and balance beam walk [126]. This suggests that, at least in mouse, ASIC3 also contributes to proprioception, albeit to a lesser degree than Piezo2.

Whether other ASIC family members also contribute to proprioceptive mechanotransduction is still unknown.

Functional ASIC channels are assembled from three homo/heterotrimeric subunits, and different combinations of ASIC subtypes display different electrophysiological properties [125,127]. However, the exact ASIC combinations in proprioceptive neurons remain unclear and need further investigation. Nonetheless, since ASIC3 was shown to be important in proprioceptive sensing, it would be interesting to examine whether it also plays a role in regulating skeletal integrity and alignment.

While the above studies have identified several ion channels that are required for proprioceptive neuron activity, the molecular signaling within sensory neurons remains elusive. To identify some of these molecules, a transcriptome analysis of proprioceptors was recently performed, revealing molecular markers for proprioceptive neurons, including specific markers for muscle spindles and GTOs [101]. Interestingly, some of these putative markers encode for ion channels and regulatory channel molecules, which are potential candidates for regulating proprioceptive signaling. We wonder whether different, perhaps more subtle, proprioceptive impairments would result in similar spine alignment and joint phenotypes, or in other musculoskeletal defects.

#### **b. Muscular components of the proprioception sensory organs**

Major components of muscle spindle are specialized muscle fibers termed intrafusal fibers. The intrafusal muscle fibers are innervated by proprioceptive sensory afferents and are morphologically distinct from their surrounding extrafusal fibers. The three types of intrafusal fibers, termed nuclear bag1, bag2 and chain fibers, differ in their myonuclear arrangement [128,129].

In muscle spindle, differentiation of intrafusal fibers begins with the connection between sensory afferents expressing neuregulin 1 (NRG1) and primary myotubes expressing its receptor ErbB2 receptor tyrosine kinase 2 (ErbB2, also known as HER2) [130–132]. NRG1–ErbB2 signaling activates downstream targets such as early growth response 3 (Egr3), which mediates the development and maturation of muscle spindle in mice [42,43,133]. Analyses of *Egr3* KO mice, which have non-functional muscle spindles, revealed skeletal malalignment [37<sup>\*\*</sup>,53<sup>\*\*</sup>,84<sup>\*\*</sup>], indicating that proprioceptive input from muscle spindles is required not only for motor coordination but also for skeletal alignment. However, the muscular components within intrafusal muscle fibers that are important for their function are still mainly unknown.

Recent advances in RNA sequencing have endorsed the analysis of isolated nuclei from muscle fibers. Recently, single-nucleus RNA sequencing of muscle spindle identified six different nuclear subtypes inside intrafusal fibers that refer to different compartments [134]. A

comparison between neuromuscular junction (NMJ) and myotendinous junction (MTJ) clusters of extrafusal and intrafusal fibers revealed high similarity between intrafusal and extrafusal muscle fibers in these zones [134]. However, it would be interesting to identify the elements in the transcriptome that separate between the two fiber types. In the context of regulation of skeletal integrity by proprioception, the association between mutations in genes identified in each cluster and skeletal abnormalities should be investigated. Notably, the bag and intrafusal-NMJ clusters were shown to express the transcript of *Piezo2*, raising the possibility that it might also function inside intrafusal fibers. However, this potential role of *Piezo2* is yet to be elucidated.

This snRNA-seq analysis has set the ground for future studies to compare the transcriptomes of intrafusal and extrafusal fibers, with aim to identify genes that are responsible for the specialized functions of each type of fibers.

#### *Myosin composition and skeletal alignment*

A major difference between intrafusal and extrafusal muscle fibers is the identity of their heavy-chain subunits of myosin (MyHCs), which include MyHC-emb (*MYH3*), MyHC-neo (*MYH8*), MyHC-slow (*MYH7*), MyHC- $\alpha$  cardiac (*MYH6*) and ancient myosins, MYH7b and MYH15 [135–139]. Moreover, it was shown that bag and chain fibers also differ in MyHC composition, as bag fibers express MyHC-slow and MyHC- $\alpha$  cardiac, and chain fibers express MyHC-emb and MyHC-neo isoforms [140]. This suggests that myosin composition may play an important role in muscle spindle function. Indeed, bag and chain fibers also differ in their functions, as bag1 fibers are solely responsible for the dynamic stretch response, whereas both bag2 and chain fibers contribute to the static stretch response.

In humans, mutations in the *MYH3* gene coding for MyHC-emb have been associated with two major distal arthrogryposis (DA) syndromes, namely Freeman–Sheldon syndrome (FSS) and Sheldon–Hall syndrome (SHS) [141]. FSS is characterized by congenital facial and limb contractures in addition to congenital scoliosis. How can embryonic myosin cause contractures of adult muscle and scoliosis? A recent study in *Myh3* mutant mice has shown the effects of MyHC-emb on myogenesis, regulating MyHC composition, fiber type, number and size [142]. These findings suggest that the contracture phenotype might be due to altered muscle size, type and MyHC composition. Additionally, adult *Myh3*-null mice exhibit scoliosis, similarly to the human FSS phenotype. Given the restricted expression of MyHC-emb to intrafusal muscle in adults, we wonder whether the contractile and scoliotic phenotypes are partly due to a nonautonomous effect of muscle spindle and proprioception on extrafusal muscles and spine. Additional studies, such

as analyzing the muscle spindle structure and function in *Myh3*-null mice and blocking *Myh3* expression specifically in muscle spindles, may reveal the involvement of proprioception in the FSS and SHS phenotypes.

*Myh3* mutation was also implicated in spondylorcarpotarsal synostosis (SCT), a skeletal disorder characterized by progressive vertebral, carpal and tarsal fusions [143]. This study identified a regulatory role for *Myh3* in the TGF $\beta$  signaling pathway, and proposed that altered signal transduction in spinal muscles may lead to the development of vertebral fusions [143]. These findings suggest a nonautonomous role for *Myh3* in skeletal regulation.

**Muscle spindle in muscular dystrophy and skeletal alignment**  
Insight into other muscular components that may affect proprioception can come from muscular diseases. Many neuromuscular diseases are accompanied by impaired function of muscle spindles, resulting in a decline of motor performance and coordination. However, in many cases, such as in Parkinson's disease and amyotrophic lateral sclerosis [144–146], proprioceptive impairment results from degeneration of neurons. Conversely, muscle diseases such as muscular dystrophy (MD), where the muscles are primarily affected, might also affect intrafusal fibers. Indeed, MD patients often suffer from balance and posture problems, suggesting that their proprioceptive system might be impaired. The most common form of MD is Duchenne muscular dystrophy (DMD), an X-linked recessive disorder caused by a mutation in the gene encoding the protein dystrophin [147]. DMD patients also develop scoliosis; however, the reason for this is poorly understood [148]. A recent study has analyzed muscle spindles from wild type and dystrophic mice, revealing a concentration of dystrophin and  $\beta$ -dystroglycan in intrafusal fibers outside the region of contact with the sensory neuron [149]. While the morphology and number of muscle spindles were similar in dystrophic and wild-type mice, an increased dynamic sensitivity of muscle spindle afferent responses was seen in dystrophic mice [149]. This altered proprioceptive response might contribute to the unstable gait and scoliosis observed in DMD patients. Again, this raises the possibility that some of the DMD skeletal phenotypes are the consequence of abnormal regulation of skeletal muscles by muscle spindle and proprioception.

### c. Capsule and extracellular matrix components

Both muscle spindle and GTOs are surrounded by two capsular structures. It has been suggested that the muscle spindle outer and inner capsules are similar to the perineurium and the endoneurial connective-tissue cells of the peripheral nerves [150–152], suggesting that they act as a selective permeable filter to control the extracellular content. The capsule is composed of connective tissue and extracellular matrix (ECM) components such as collagens, laminins, heparan sulfate and fibronectins [153,154]. However, the development and function of the capsule is still

largely unknown. To our knowledge, the question of whether the capsule plays a role in muscle spindle and GTO development and function has not been addressed.

Since the capsule is enriched with ECM molecules, we wonder whether ECM-related myopathy phenotypes are related to impaired proprioception. For example, the outer capsule, containing a basement membrane, was shown to express collagen type VI (colVI) [154]. ColVI is associated with two main types of muscle disorders: Bethlem myopathy (BM) and the more severe syndrome Ullrich congenital muscular dystrophy (UCMD) [155,156]. Typically, UCMD patients present muscle weakness and hypotonia, congenital hip dysplasia, and contractures including torticollis, kyphoscoliosis and rigid spine [156]. A mouse model of colVI deficiency has been generated by deletion of the *Col6a1* gene [157]. These mice exhibit histological features of myopathy, such as fiber necrosis and variation in fiber width, but without obvious weakness. Subsequent work using these mice linked colVI functionally also to the peripheral nervous system, demonstrating that lack of colVI leads to increased myelin thickness, impaired nerve conduction, impaired motor coordination and delayed response to acute pain stimuli [158]. This raises the hypothesis that some of the UCMD phenotypes arise from a nonautonomous effect of ECM component in glia and perhaps also in proprioception tissues on the musculoskeletal system. To examine this hypothesis, a capsule-specific deletion of colIV should be studied.

### Conclusions and future directions

In this review, we focused on the contribution of the proprioceptive system to musculoskeletal biology. We have described the non-traditional roles of proprioception in spinal alignment, realignment of fractured bones and joint morphogenesis (summarized in Figure 1). Interestingly, in three different genetic mouse models, impaired proprioceptive function leads to the same skeletal deformations, suggesting non-autonomous role for the proprioceptive system in skeletal biology. Furthermore, conditional deletion of *Piezo2* in proprioceptive neurons, but not in skeletal tissues, resulted in these skeletal deformation, further supporting this non-autonomous role. However, we believe that these skeletal phenotypes are only the tip of the iceberg and that future work will identify additional aspects of musculoskeletal biology that are regulated by the proprioceptive system.

A key question that remains open relates to the mechanisms underlying the regulation of musculoskeletal development by proprioceptive signaling. We speculate that different but nonexclusive mechanisms may be involved. For example, in the case of spine alignment, it is possible that the regulatory mechanism involves muscle tensions from both sides of the spine. Thus, loss of proprioception may lead to abnormal muscle tensions, resulting in spine misalignment. In joints, loss of

proprioception leads to uncoordinated movement and muscle activation patterns, which can lead to abnormal stresses on the joint. Such abnormal stresses can lead to abnormal mechanical signals in joint cells, affecting joint integrity and resulting in aberrant joint morphology. However, further studies are needed to verify these hypotheses and to explain the regulatory role of proprioception in skeletal biology.

We argue that the proprioceptive system regulates numerous aspects of musculoskeletal development. However, the effect of the proprioception system on the development of non-skeletal tissues has yet to be established. It remains to be explored how the proprioceptive system affects muscle function, diseases and repair. Additionally, recent studies have shown that in different myopathies, the intrafusal fibers are also impaired, raising the possibility that aberrant proprioceptive function is involved in the pathogenesis of these diseases.

Finally, there is still a need to identify the molecular players that mediate proprioceptive sensing. While the molecular apparatus of proprioceptive neurons is beginning to emerge, the identification of muscular and capsular components of proprioception has lagged behind. This gap in molecular knowledge prevents a comprehensive understanding of proprioceptor development and function. Moreover, it prevents associating molecular information on human musculoskeletal pathologies with proprioceptive function. Thus, an effort should be made to reveal the complete molecular identity of proprioceptors.

The involvement of the proprioceptive system in development, maintenance and repair of the skeleton increases substantially the scope of known functions of this system. Moreover, it raises the possibility that the proprioceptive system is involved in regulating other processes and that its dysfunction may contribute to the etiology of various musculoskeletal pathologies. Thus, further research of this fascinating system may have important medical implications.

### Conflict of interest statement

Nothing declared.

### Acknowledgements

This work was supported by the Cottrel Foundation, the Estate of Mr. and Mrs. van Adelsbergen, the David and Fela Shapell Family Center for Genetic Disorders, and by Eric and Julie Borman (to E.Z.), and by N.I.H. (grant number NS086973) and NSF (grant number 2015317) (to M.C.T).

### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Proske U, Gandevia SC: **The proprioceptive senses: their roles in signaling body shape, body position and movement, and muscle force.** *Physiol Rev* 2012, **92**:1651-1697.
2. Dietz V: **Proprioception and locomotor disorders.** *Nat Rev Neurosci* 2002, **3**:781-790.
3. Bewick GS, Banks RW: **Mechanotransduction in the muscle spindle.** *Pflugers Arch Eur J Physiol* 2014, **467**:175-190.
4. Jami L: **Golgi tendon organs in mammalian skeletal muscle: functional properties and central actions.** *Physiol Rev* 1992, **72**:623-666.
5. Moore JC: **The Golgi tendon organ: a review and update.** *Am J Occup Ther Off Publ Am Occup Ther Assoc* 1984, **38**:227-236.
6. Chen HH, Hippenmeyer S, Arber S, Frank E: **Development of the monosynaptic stretch reflex circuit.** *Curr Opin Neurobiol* 2003, **13**:96-102.
7. Chen HH, Frank E: **Development and specification of muscle sensory neurons.** *Curr Opin Neurobiol* 1999, **9**:405-409.
8. Shwartz Y, Blitz E, Zelzer E: **One load to rule them all: mechanical control of the musculoskeletal system in development and aging.** *Differentiation* 2013, **86**:104-111.
9. Felsenthal N, Zelzer E: **Mechanical regulation of musculoskeletal system development.** *Dev* 2017, **144**:4271-4283.
10. Reamy BV, Slakey JB: **Adolescent idiopathic scoliosis: review and current concepts.** *Am Fam Physician* 2001.
11. Kouwenhoven JWM, Castelein RM: **The pathogenesis of adolescent idiopathic scoliosis: review of the literature.** *Spine (Phila Pa 1976)* 2008, **33**:2898-2908.
12. Ouellet J, Odent T: **Animal models for scoliosis research: state of the art, current concepts and future perspective applications.** *Eur Spine J* 2013, **22**:81-95.
13. Shi L, Wang D, Hui SCN, Tong MCF, Cheng JCY, Chu WCW: **Volumetric changes in cerebellar regions in adolescent idiopathic scoliosis compared with healthy controls.** *Spine J* 2013, **13**:1904-1911.
14. Wang D, Shi L, Chu WCW, Burwell RG, Cheng JCY, Ahuja AT: **Abnormal cerebral cortical thinning pattern in adolescent girls with idiopathic scoliosis.** *Neuroimage* 2012, **59**:935-942.
15. Shi L, Wang D, Chu WCW, Burwell GR, Wong TT, Heng PA, Cheng JCY: **Automatic MRI segmentation and morphoanatomy analysis of the vestibular system in adolescent idiopathic scoliosis.** *Neuroimage* 2011, **54**(Suppl 1): S180-S188.
16. Guo X, Chau WW, Hui-Chan CWY, Cheung CSK, Tsang WWN, Cheng JCY: **Balance control in adolescents with idiopathic scoliosis and disturbed somatosensory function.** *Spine (Phila Pa 1976)* 2006, **31**:E437-E440.
17. Acaroglu E, Akel I, Alanay A, Yazici M, Marcucio R: **Comparison of the melatonin and calmodulin in paravertebral muscle and platelets of patients with or without adolescent idiopathic scoliosis.** *Spine (Phila Pa 1976)* 2009, **34**:E659-E663.
18. McIntire KL, Asher MA, Burton DC, Liu W: **Trunk rotational strength asymmetry in adolescents with idiopathic scoliosis: an observational study.** *Scoliosis* 2007, **2**:1-9.
19. Gillen G: *Stroke Rehabilitation*. 2015.
20. Smorenburg ARP, Ledebt A, Deconinck FJA, Savelsbergh GJP: **Deficits in upper limb position sense of children with Spastic Hemiparetic Cerebral Palsy are distance-dependent.** *Res Dev Disabil* 2012, **33**:971-981.
21. Barrios C, Tuñón MT, De salis JA, Beguiristain JL, Cañadell J: **Scoliosis induced by medullary damage: an experimental study in rabbits.** *Spine (Phila Pa 1976)* 1987, **12**:433-439.
22. Pincott JR, Davies JS, Taffs LF: **Scoliosis caused by section of dorsal spinal nerve roots.** *J Bone Jt Surg - Ser B* 1984, **66**:27-29.
23. MacEwen GD: **Experimental scoliosis.** *Isr J Med Sci* 1973, **9**:714-718.
24. Dukelow SP, Herter TM, Moore KD, Demers MJ, Glasgow JI, Bagg SD, Norman KE, Scott SH: **Quantitative assessment of**

- limb position sense following stroke. *Neurorehabil Neural Repair* 2010, **24**:178-187.**
25. Smith DL, Akhtar AJ, Garraway WM: **Proprioception and spatial neglect after stroke.** *Age Ageing* 1983, **12**:63-69.
  26. Yang JH, Suh SW, Sung PS, Park WH: **Asymmetrical gait in adolescents with idiopathic scoliosis.** *Eur Spine J* 2013, **22**:2407-2413.
  27. Gruber AH, Busa MA, Gorton GE, Van Emmerik REA, Masso PD, Hamill J: **Time-to-contact and multiscale entropy identify differences in postural control in adolescent idiopathic scoliosis.** *Gait Posture* 2011, **34**:13-18.
  28. Lao MLM, Chow DHK, Guo X, Cheng JCY, Holmes AD: **Impaired dynamic balance control in adolescents with idiopathic scoliosis and abnormal somatosensory evoked potentials.** *J Pediatr Orthop* 2008, **28**:846-849.
  29. Barrack RL, Whitecloud TS, Burke SW, Cook SD, Harding AF: **Proprioception in idiopathic scoliosis.** *Spine (Phila Pa 1976)* 1984, **9**:681-685.
  30. Wyatt MP, Barrack RL, Mubarak SJ, Whitecloud TS, Burke SW: **Vibratory response in idiopathic scoliosis.** *J Bone Jt Surg - Ser B* 1986, **68**:714-718.
  31. Yekutieli M, Robin GC, Yarom R: **Proprioceptive function in children with adolescent idiopathic scoliosis.** *Spine (Phila Pa 1976)* 1981, **6**:560-566.
  32. Ford DM, Bagnall KM, Clements CA, McFadden KD: **Muscle spindles in the paraspinal musculature of patients with adolescent idiopathic scoliosis.** *Spine (Phila Pa 1976)* 1988, **13**:461-465.
  33. Goble DJ, Lewis CA, Hurvitz EA, Brown SH: **Development of upper limb proprioceptive accuracy in children and adolescents.** *Hum Mov Sci* 2005, **24**:155-170.
  34. Pickett K, Konczak J: **Measuring kinaesthetic sensitivity in typically developing children.** *Dev Med Child Neurol* 2009, **51**:711-716.
  35. Levanon D, Bettoun D, Harris-Cerruti C, Woolf E, Negreanu V, Eilam R, Bernstein Y, Goldenberg D, Xiao C, Fliegauf M *et al.*: **The Runx3 transcription factor regulates development and survival of TrkC dorsal root ganglia neurons.** *EMBO J* 2002, **21**:3454-3463.
  36. Inoue ichi K, Ozaki S, Shiga T, Ito K, Masuda T, Okado N, Iseda T, Kawaguchi S, Ogawa M, Bae SC *et al.*: **Runx3 controls the axonal projection of proprioceptive dorsal root ganglion neurons.** *Nat Neurosci* 2002, **5**:946-954.
  37. Blecher R, Krief S, Gallili T, Biton IE, Stern T, Assaraf E, Levanon D, Appel E, Anekstein Y, Agar G *et al.*: **The proprioceptive system masterminds spinal alignment: insight into the mechanism of scoliosis.** *Dev Cell* 2017, **399**:388-399.e3
- This work used genetic mouse models to demonstrate the involvement of proprioception in regulating spine alignment. The authors show that *Runx3* null mice, which lack functional proprioceptive neurons, develop peripubertal scoliosis without prior vertebral dysplasia or muscle asymmetry. A similar phenotype but of reduced severity was seen in *Egr3* null mice, which lack muscle spindles but not Golgi tendon organs. Functional assays revealed a decrease in gait regularity, which was also more pronounced in *Runx3* null mice than in *Egr3* null mice. These findings implicate impaired proprioceptive signaling in acquired scoliosis and suggest that both receptor types are required for this regulatory mechanism.
38. Stricker S, Fundele R, Vortkamp A, Mundlos S: **Role of Runx genes in chondrocyte differentiation.** *Dev Biol* 2002, **245**:95-108.
  39. Yoshida CA, Yamamoto H, Fujita T, Furuichi T, Ito K, Inoue KI, Yamana K, Zanma A, Takada K, Ito Y *et al.*: **Runx2 and Runx3 are essential for chondrocyte maturation, and Runx2 regulates limb growth through induction of Indian hedgehog.** *Genes Dev* 2004, **18**:952-963.
  40. Bauer O, Sharir A, Kimura A, Hantisteanu S, Takeda S, Groner Y: **Loss of osteoblast Runx3 produces severe congenital osteopenia.** *Mol Cell Biol* 2015, **35**:1097-1109.
  41. Lian JB, Javed A, Zaidi SK, Lengner C, Montecino M, Van Wijnen AJ, Stein JL, Stein GS: **Regulatory controls for osteoblast growth and differentiation: role of Runx/Cbfa/AML factors.** *Crit Rev Eukaryot Gene Expr* 2004, **14**:1-41.
  42. Tourtellotte WG, Keller-Peck C, Milbrandt J, Kucera J: **The transcription factor Egr3 modulates sensory axon-myotube interactions during muscle spindle morphogenesis.** *Dev Biol* 2001, **232**:388-399.
  43. Tourtellotte WG, Milbrandt J: **Sensory ataxia and muscle spindle agenesis in mice lacking the transcription factor Egr3.** *Nat Genet* 1998, **20**:87-91.
  44. Ellsasser JC, Moyer CF, Lesker PA, Simmons DJ: **Improved healing of experimental long bone fractures in rabbits by delayed internal fixation.** *J Trauma - Inj Infect Crit Care* 1975, **15**:869-876.
  45. Fogel GR, Morrey BF: **Delayed open reduction and fixation of ankle fractures.** *Clin Orthop Relat Res* 1987, **215**:187-195.
  46. Court-Brown CM: **Principles of nonoperative fracture treatment.** *Rockwood, Green, and Wilkins Fractures in Adults and Children.* edn 8. 2014:699-707.
  47. Husain SN, King EC, Young JL, Sarwark JF: **Remodeling of birth fractures of the humeral diaphysis.** *J Pediatr Orthop* 2008, **28**:10-13.
  48. Bramblett CA: **Pathology in the Darajani baboon.** *Am J Phys Anthropol* 1967, **26**:331-340.
  49. Duckworth WL: **On the Natural Repair of Fractures, as seen in the Skeletons of Anthropoid Apes.** *J Anat* 1911, **46**:81-85.
  50. Schultz AH: **Age changes and variability in gibbons.** *A Morphological study on a population sample of a man-like ape.* 1944.
  51. Schultz JD: *Bones: Structure and mechanics.* 2013.
  52. Rot C, Stern T, Blecher R, Friesem B, Zelzer E: **A mechanical jack-like mechanism drives spontaneous fracture healing in neonatal mice.** *Dev Cell* 2014, **31**:159-170.
  53. Blecher R, Krief S, Gallili T, Assaraf E, Stern T, Anekstein Y, Agar G, Zelzer E: **The proprioceptive system regulates morphologic restoration of fractured bones.** *Cell Rep* 2017, **20**:1775-1783
- This work used genetic mouse models to demonstrate the involvement of proprioception in regulating the realignment of fractured bones, termed natural reduction. In *Runx3* null mice, which lack functional proprioceptive neurons, and in conditional knockout mice lacking *Runx3* in the peripheral nervous system, natural reduction failed to occur. The process failed also upon inactivation of muscles around the fracture site. A similar but less severe phenotype was seen in *Egr3*, which lack muscle spindles but not Golgi tendon organs. These findings suggest that both proprioceptor types, as well as muscle contraction, are necessary for the regulation of natural reduction.
54. Zelzer E, Blitz E, Killian ML, Thomopoulos S: **Tendon-to-bone attachment: from development to maturity.** *Birth Defects Res Part C - Embryo Today Rev* 2014, **102**:101-112.
  55. Brotto M, Johnson ML: **Endocrine crosstalk between muscle and bone.** *Curr Osteoporos Rep* 2014, **12**:135-141.
  56. Colaianni G, Storlino G, Sanesi L, Colucci S, Grano M: **Myokines and osteokines in the pathogenesis of muscle and bone diseases.** *Curr Osteoporos Rep* 2020, **18**:401-407.
  57. Kirk B, Feehan J, Lombardi G, Duque G: **Muscle, bone, and fat crosstalk: the biological role of myokines, osteokines, and adipokines.** *Curr Osteoporos Rep* 2020, **18**:388-400.
  58. Sharir A, Stern T, Rot C, Shahar R, Zelzer E: **Muscle force regulates bone shaping for optimal loadbearing capacity during embryogenesis.** *Development* 2011, **138**:3247-3259.
  59. Kahn J, Shwartz Y, Blitz E, Krief S, Sharir A, Breitel DA, Rattenbach R, Relaix F, Maire P, Rountree RB *et al.*: **Muscle contraction is necessary to maintain joint progenitor cell fate.** *Dev Cell* 2009, **16**:734-743.
  60. Abou-Khalil R, Yang F, Lieu S, Julien A, Perry J, Pereira C, Relaix F, Miclau T, Marcucio R, Colnot C: **Role of muscle stem cells during skeletal regeneration.** *Stem Cells* 2015, **33**:1501-1511.

61. Currey JD: **The many adaptations of bone.** *J Biomech* 2003, **36**:1487-1495.
62. Frost HM: **From Wolff's law to the Utah paradigm: insights about bone physiology and its clinical applications.** *Anat Rec* 2001, **262**:398-419.
63. Weiner S, Wagner HD: **The material bone: structure-mechanical function relations.** *Annu Rev Mater Sci* 1998, **28**:271-298.
64. Burr DB, Allen MR: *Basic and Applied Bone Biology*. 2013.
65. Robling AG, Castillo AB, Turner CH: **Biomechanical and molecular regulation of bone remodeling.** *Annu Rev Biomed Eng* 2006, **8**:455-498.
66. Bach-Gansmo FL, Wittig NK, Brüel A, Thomsen JS, Birkedal H: **Immobilization and long-term recovery results in large changes in bone structure and strength but no corresponding alterations of osteocyte lacunar properties.** *Bone* 2016, **91**:139-147.
67. Ellman R, Spatz J, Cloutier A, Palme R, Christiansen BA, Bouxsein ML: **Partial reductions in mechanical loading yield proportional changes in bone density, bone architecture, and muscle mass.** *J Bone Miner Res* 2013, **28**:875-885.
68. Davidson RM, Tatakis DW, Auerbach AL: **Multiple forms of mechanosensitive ion channels in osteoblast-like cells.** *Pflügers Arch Eur J Physiol* 1990, **416**:646-651.
69. Huiskes R, Rulmerman R, Van Lenthe GH, Janssen JD: **Effects of mechanical forces on maintenance and adaptation of form in trabecular bone.** *Nature* 2000, **405**:704-706.
70. Lee HS, Millward-Sadler SJ, Wright MO, Nuki G, Salter DM: **Integrin and mechanosensitive ion channel-dependent tyrosine phosphorylation of focal adhesion proteins and  $\beta$ -catenin in human articular chondrocytes after mechanical stimulation.** *J Bone Miner Res* 2000, **15**:1501-1509.
71. Wann AKT, Zuo N, Haycraft CJ, Jensen CG, Poole CA, McGlashan SR, Knight MM: **Primary cilia mediate mechanotransduction through control of ATP-induced Ca<sup>2+</sup> signaling in compressed chondrocytes.** *FASEB J* 2012, **26**:1663-1671.
72. Aro HT, Chao EYS: **Bone-healing patterns affected by loading, fracture fragment stability, fracture type, and fracture site compression.** *Clinical Orthopaedics and Related Research*. 1993:8-17.
73. Probst A, Spiegel HU: **Cellular mechanisms of bone repair.** *J Investig Surg* 1997, **10**:77-86.
74. Thompson Z, Miclau T, Hu D, Helms JA: **A model for intramembranous ossification during fracture healing.** *J Orthop Res* 2002, **20**:1091-1098.
75. Maeda N, Osawa K, Masuda T, Hakeda Y, Kumegawa M: **Postnatal development of the annulospiral endings of Ia fibers in muscle spindles of mice.** *Acta Anat (Basel)* 1985, **124**:42-46.
76. Osawa K, Maeda N, Sato M, Kawasaki T, Masuda T, Yamamoto Y, Hakeda Y, Ukai M, Watanabe Y, Suwa T et al.: **Postnatal development of the annulospiral endings of Ia fibers in muscle spindles of the mouse temporal muscle.** *Anat Anz* 1988, **167**:253-257.
77. Noordin S, Umer M, Hafeez KNH: **Developmental dysplasia of the hip.** *Orthop Rev* 2010, **2**:e19.
78. Vaquero-Picado A, González- Morán G, Garay EG, Moraleda L: **Developmental dysplasia of the hip: update of management.** *EFORT Open Rev* 2019, **4**:548-556.
79. Guille JT, Pizzutillo PDMG: **Development dysplasia of the hip from birth to six months.** *J Am Acad Orthop Surg* 2000, **8**:232-242.
80. Wynne-Davies R: **A family study of neonatal and late-diagnosis congenital dislocation of the hip.** *J Med Genet* 1970, **7**:315-333.
81. Kotlarsky P, Haber R, Bialik V, Eidelman M: **Developmental dysplasia of the hip: what has changed in the last 20 years?** *World J Orthop* 2015 <http://dx.doi.org/10.5312/wjo.v6.i11.886>.
82. Levanon D, Groner Y: **Runx3-deficient mouse strains circa 2008: resemblance and dissimilarity.** *Blood Cells Mol Dis* 2009, **43**:1-15.
83. Woo SH, Lukacs V, De Nooij JC, Zaytseva D, Criddle CR, Francisco A, Jessell TM, Wilkinson KA, Patapoutian A: **Piezo2 is the principal mechanotransduction channel for proprioception.** *Nat Neurosci* 2015, **18**:1756-1762.
84. Assaraf E, Blecher R, Heinemann-Yerushalmi L, Krief S, Carmel •• Vinestock R, Biton IE, Brumfeld V, Rotkopf R, Avisar E, Agar G et al.: **Piezo2 expressed in proprioceptive neurons is essential for skeletal integrity.** *Nat Commun* 2020, **11**:3168
- This work shows that loss of *Piezo2* in mice recapitulates several human skeletal abnormalities, such as scoliosis and hip dysplasia. Conditional knockout of *Piezo2* in proprioceptive neurons, chondrogenic or osteogenic lineages indicated that it functions specifically in proprioceptive neurons to regulate spine alignment and hip morphogenesis. The non-autonomous role of proprioception in hip joint morphogenesis was further validated in mice mutant for proprioceptive system regulators *Runx3* or *Egr3*, which displayed similar joint abnormalities. These findings expand the range of known regulatory roles of the proprioception system on the skeleton and provide a central component of the underlying molecular mechanism, namely *Piezo2*.
85. Farrokhi S, Voycheck CA, Tashman S, Fitzgerald GK: **A biomechanical perspective on physical therapy management of knee osteoarthritis.** *J Orthop Sports Phys Ther* 2013, **43**:600-619.
86. Felson DT: **Osteoarthritis as a disease of mechanics.** *Osteoarthr Cartil* 2013, **21**:10-15.
87. Solomonow M: **Sensory - Motor control of ligaments and associated neuromuscular disorders.** *J Electromyogr Kinesiol* 2006, **16**:549-567.
88. Pietrosimone BG, McLeod MM, Lepley AS: **A theoretical framework for understanding neuromuscular response to lower extremity joint injury.** *Sports Health* 2012, **4**:31-35.
89. Herzog W, Longino D, Clark A: **The role of muscles in joint adaptation and degeneration.** *Langenbeck's Archives of Surgery*. 2003.
90. Sandercock TG, Wei Q, Dhaher YY, Pai DK, Tresch MC: **Vastus lateralis and vastus medialis produce distinct mediolateral forces on the patella but similar forces on the tibia in the rat.** *J Biomech* 2018, **81**:45-51.
91. Alessandro C, Barroso FO, Prashara A, Tentler DP, Yeh HY, Tresch MC: **Coordination amongst quadriceps muscles suggests neural regulation of internal joint stresses, not simplification of task performance.** *Proc Natl Acad Sci U S A* 2020, **117**:8135-8142.
92. Alessandro C, Rellinger BA, Barroso FO, Tresch MC: **Adaptation after vastus lateralis denervation in rats demonstrates neural regulation of joint stresses and strains.** *Elife* 2018, **7**:e38215
- This study tested the hypothesis that the central nervous system controls muscle activation to avoid excessive joint stresses and strains. Muscle activation strategies were examined after selective paralysis of a muscle acting on the rat's knee, showing compromises between restoration of task performance and regulation of joint stresses and strains. These results suggest that the nervous system induces specific muscle activation patterns that minimize joint stresses and strains, while achieving desired task performance.
93. Barroso FO, Alessandro C, Tresch MC: **Adaptation of muscle activation after patellar loading demonstrates neural control of joint variables.** *Sci Rep* 2019, **9**:20370
- This study tested the hypothesis that the central nervous system controls muscle activation to avoid excessive joint stresses and strains. The authors analyzed the coordination between rat quadriceps muscles before and after imposing a lateral force on the patella by attaching a spring between the patella and lateral femur. A few days later, an adaptation of the ratio between activations of the various muscles was observed, which was reversed upon removal of the spring. These results are consistent with the hypothesis that the nervous system chooses muscle activations to control joint stress.
94. Roos EM, Herzog W, Block JA, Bennell KL: **Muscle weakness, afferent sensory dysfunction and exercise in knee osteoarthritis.** *Nat Rev Rheumatol* 2011, **7**:57-63.

95. O'Connor BL, Visco DM, Brandt KD, Albrecht M, O'Connor AB: **Sensory nerves only temporarily protect the unstable canine knee joint from osteoarthritis. evidence that sensory nerves reprogram the central nervous system after cruciate ligament transection.** *rthritis Rheum* 1993, **36**:1154-1163.
96. Grigg P, Greenspan BJ: **Response of primate joint afferent neurons to mechanical stimulation of knee joint.** *J Neurophysiol* 1977, **40**:1-8.
97. Wilmink RJH, Nichols TR: **Distribution of heterogenic reflexes among the quadriceps and triceps surae muscles of the cat hind limb.** *J Neurophysiol* 2003, **90**:2310-2324.
98. Roy RR, Kim JA, Monti RJ, Zhong H, Edgerton VR: **Architectural and histochemical properties of cat hip 'Cuff' muscles.** *Cells Tissues Organs* 1997, **159**:136-146.
99. Maass S, Baumann KI, Halata Z: **Topography of muscle spindles and Golgi tendon organs in shoulder muscles of "monodelphis domestica."** *Ann Anat* 2001, **183**:237-242.
100. Riemann BL, Lephart SM: **The sensorimotor system, Part II: the role of proprioception in motor control and functional joint stability.** *J Athl Train* 2002, **37**:80-84.
101. Wu D, Schieren I, Qian Y, Zhang C, Jessell TM, de Nooij JC: **A role for sensory end organ-derived signals in regulating muscle spindle proprioceptor phenotype.** *J Neurosci* 2019, **39**:4252-4267.
102. Faure L, Wang Y, Kastriti ME, Fontanet P, Cheung KKY, Petitpré C, Wu H, Sun LL, Runge K, Croci L *et al.*: **Single cell RNA sequencing identifies early diversity of sensory neurons forming via bi-potential intermediates.** *Nat Commun* 2020 <http://dx.doi.org/10.1038/s41467-020-17929-4>.
103. Zheng Y, Liu P, Bai L, Trimmer JS, Bean BP, Ginty DD: **Deep sequencing of somatosensory neurons reveals molecular determinants of intrinsic physiological properties.** *Neuron* 2019, **103**:598-616.e7.
104. Saotome K, Murthy SE, Kefauver JM, Whitwam T, Patapoutian A, Ward AB: **Structure of the mechanically activated ion channel Piezo1.** *Nature* 2018, **554**:481-486.
105. Zhao Q, Zhou H, Chi S, Wang Y, Wang J, Geng J, Wu K, Liu W, Zhang T, Dong MQ *et al.*: **Structure and mechanogating mechanism of the Piezo1 channel.** *Nature* 2018, **554**:487-492.
106. Wang L, Zhou H, Zhang M, Liu W, Deng T, Zhao Q, Li Y, Lei J, Li X, Xiao B: **Structure and mechanogating of the mammalian tactile channel PIEZO2.** *Nature* 2019, **573**:225-229.
107. Murthy SE, Dubin AE, Patapoutian A: **Piezos thrive under pressure: mechanically activated ion channels in health and disease.** *Nat Rev Mol Cell Biol* 2017, **18**:771-783.
108. Nonomura K, Woo SH, Chang RB, Gillich A, Qiu Z, Francisco AG, Ranade SS, Liberles SD, Patapoutian A: **Piezo2 senses airway stretch and mediates lung inflation-induced apnoea.** *Nature* 2017, **541**:176-181.
109. Zhong M, Komarova Y, Rehman J, Malik AB: **Mechanosensing Piezo channels in tissue homeostasis including their role in lungs.** *Pulm Circ* 2018, **8**:1-6.
110. Wang F, Knutson K, Alcaino C, Linden DR, Gibbons SJ, Kashyap P, Grover M, Oeckler R, Gottlieb PA, Li HJ *et al.*: **Mechanosensitive ion channel Piezo2 is important for enterochromaffin cell response to mechanical forces.** *J Physiol* 2017, **595**:79-91.
111. Woo SH, Ranade S, Weyer AD, Dubin AE, Baba Y, Qiu Z, Petrus M, Miyamoto T, Reddy K, Lumpkin EA *et al.*: **Piezo2 is required for Merkel-cell mechanotransduction.** *Nature* 2014, **509**:622-626.
112. Lee W, Leddy HA, Chen Y, Lee SH, Zelenski NA, McNulty AL, Wu J, Beicker KN, Coles J, Zauscher S *et al.*: **Synergy between Piezo1 and Piezo2 channels confers high-strain mechanosensitivity to articular cartilage.** *Proc Natl Acad Sci U S A* 2014, **111**:E5114-E5122.
113. Mahmud AA, Nahid NA, Nassif C, Sayeed MSB, Ahmed MU, Parveen M, Khalil MI, Islam MM, Nahar Z, Rypens F *et al.*: **Loss of the proprioception and touch sensation channel PIEZO2 in siblings with a progressive form of contractures.** *Clin Genet* 2017, **91**:470-475.
114. Delle Vedove A, Storbeck M, Heller R, Hölker I, Hebbar M, Shukla A, Magnusson O, Cirak S, Girisha KM, O'Driscoll M *et al.*: **Biallelic loss of proprioception-related PIEZO2 causes muscular atrophy with perinatal respiratory distress, arthrogyposis, and scoliosis.** *Am J Hum Genet* 2016, **99**:1206-1216.
115. Haliloglu G, Becker K, Temucin C, Talim B, Küçüksa?hin N, Pergande M, Motameny S, Nürnberg P, Aydingoz U, Topaloglu H *et al.*: **Recessive PIEZO2 stop mutation causes distal arthrogyposis with distal muscle weakness, scoliosis and proprioception defects.** *J Hum Genet* 2017, **62**:497-501.
116. Chesler AT, Szczot M, Bharucha-Goebel D, Āeko M, Donkervoort S, Laubacher C, Hayes LH, Alter K, Zampieri C, Stanley C, Innes AM, Mah JK, Grosman CM, Bradley N, Nguyen D, Foley AR, Le Pichon CE, Bönnemann CG: **The role of PIEZO2 in human mechanosensation.** *N Engl J Med* 2016, **375**:1355-1364.
117. Zhou T, Gao B, Fan Y, Liu Y, Feng S, Cong Q, Zhang X, Zhou Y, Yadav PS, Lin J *et al.*: **Piezo1/2 mediate mechanotransduction essential for bone formation through concerted activation of NFAT-YAP1-β-catenin.** *Elife* 2020, **9**:1-38.
118. Narayanan P, Sondermann J, Rouwette T, Karaca S, Urlaub H, Mitkovski M, Gomez-Varela D, Schmidt M: **Native Piezo2 interactomics identifies pericentrin as a novel regulator of Piezo2 in somatosensory neurons.** *J Proteome Res* 2016, **15**:2676-2687.
119. Narayanan P, Hütte M, Kudryasheva G, Taberner FJ, Lechner SG, Rehfeldt F, Gomez-Varela D, Schmidt M: **Myotubularin related protein-2 and its phospholipid substrate PIP 2 control Piezo2-mediated mechanotransduction in peripheral sensory neurons.** *Elife* 2018, **7**:1-28.
120. Bolino A, Muglia M, Conforti FL, LeGuern E, Salih MAM, Georgiou DM, Christodoulou K, Hausmanowa-Petrusewicz I, Mandich P, Schenone A *et al.*: **Charcot-Marie-Tooth type 4B is caused by mutations in the gene encoding myotubularin-related protein-2.** *Nat Genet* 2000, **25**:17-19.
121. Halperin D, Sapir A, Wormser O, Drabkin M, Yogev Y, Dolgin V, Flusser H, Birk OS: **Novel MTMR2 mutation causing severe Charcot-Marie-Tooth type 4B1 disease: a case report.** *Neurogenetics* 2020, **21**:301-304.
122. Anderson EO, Schneider ER, Matson JD, Gracheva EO, Bagriantsev SN: **TMEM150C/Tentonin3 is a regulator of mechano-gated ion channels.** *Cell Rep* 2018, **23**:701-708.
123. Hong GS, Lee B, Wee J, Chun H, Kim H, Jung J, Cha JY, Riew TR, Kim GH, Kim IB *et al.*: **Tentonin 3/TMEM150c confers distinct mechanosensitive currents in dorsal-root ganglion neurons with proprioceptive function.** *Neuron* 2016, **91**:107-118.
124. Simon A, Shenton F, Hunter I, Banks RW, Bewick GS: **Amiloride-sensitive channels are a major contributor to mechanotransduction in mammalian muscle spindles.** *J Physiol* 2010, **588**:171-185.
125. Cheng YR, Jiang BY, Chen CC: **Acid-sensing ion channels: dual function proteins for chemo-sensing and mechano-sensing.** *J Biomed Sci* 2018, **25**:1-14.
126. Lin SH, Cheng YR, Banks RW, Min MY, Bewick GS, Chen CC: **Evidence for the involvement of ASIC3 in sensory mechanotransduction in proprioceptors.** *Nat Commun* 2016, **7**:11460.
127. Jasti J, Furukawa H, Gonzales EB, Gouaux E: **Structure of acid-sensing ion channel 1 at 1.9 Å resolution and low pH.** *Nature* 2007, **449**:316-323.
128. Ovalle WK, Smith RS: **Histochemical identification of three types of intrafusal muscle fibers in the cat and monkey based on the myosin ATPase reaction.** *Can J Physiol Pharmacol* 1972, **50**:195-202.
129. Hunt CC: **Mammalian muscle spindle: peripheral mechanisms.** *Physiol Rev* 1990, **7**:643-663.

130. Leu M, Bellmunt E, Schwander M, Fariñas I, Brenner HR, Müller U: **ErbB2 regulates neuromuscular synapse formation and is essential for muscle spindle development.** *Development* 2003, **130**:2291-2301.
131. Hippenmeyer S, Shneider NA, Birchmeier C, Burden SJ, Jessell TM, Arber S: **A role for Neuregulin1 signaling in muscle spindle differentiation.** *Neuron* 2002, **36**:1035-1049.
132. Cheret C, Willem M, Fricker FR, Wende H, Wulf-Goldenberg A, Tahirovic S, Nave KA, Saftig P, Haass C, Garratt AN et al.: **Bace1 and Neuregulin-1 cooperate to control formation and maintenance of muscle spindles.** *EMBO J* 2013, **32**:2015-2028.
133. Fernandes MO, Tourtellotte WG: **Egr3-dependent muscle spindle stretch receptor intrafusal muscle fiber differentiation and fusimotor innervation homeostasis.** *J Neurosci* 2015, **35**:5566-5578.
134. Kim M, Franke V, Brandt B, Lowenstein ED, Schöwel V, Spuler S, Akalin A, Birchmeier C: **Single-nucleus transcriptomics reveals functional compartmentalization in syncytial skeletal muscle cells.** *Nat Commun* 2020, **11**:1-14.
135. Walro JM, Kucera J: **Why adult mammalian intrafusal and extrafusal fibers contain different myosin heavy-chain isoforms.** *Trends Neurosci* 1999, **22**:180-184.
136. Wang J, McWhorter DL, Walro JM: **Stability of myosin heavy chain isoforms in selectively denervated: Adult rat muscle spindles.** *Anat Rec* 1997, **249**:32-43.
137. Soukup T, Pedrosa-Domellöf F, Thornell L-E: **Expression of myosin heavy chain isoforms and myogenesis of intrafusal fibres in rat muscle spindles.** *Microsc Res Tech* 1995, **30**:390-407.
138. Schiaffino S, Rossi AC, Smerdu V, Leinwand LA, Reggiani C: **Developmental myosins: Expression patterns and functional significance.** *Skelet Muscle* 2015, **5**:1-14.
139. Rossi AC, Mammucari C, Argentini C, Reggiani C, Schiaffino S: **Two novel/ancient myosins in mammalian skeletal muscles: MYH14/7b and MYH15 are expressed in extraocular muscles and muscle spindles.** *J Physiol* 2010, **588**:353-364.
140. Pedrosa F, Butler-Browne GS, Dhoot GK, Fischman DA, Thornell LE: **Diversity in expression of myosin heavy chain isoforms and M-band proteins in rat muscle spindles.** *Histochemistry* 1989, **92**:185-194.
141. Toydemir RM, Rutherford A, Whitby FG, Jorde LB, Carey JC, Bamshad MJ: **Mutations in embryonic myosin heavy chain (MYH3) cause Freeman-Sheldon syndrome and Sheldon-Hall syndrome.** *Nat Genet* 2006, **38**:561-565.
142. Agarwal M, Sharma A, Kumar P, Kumar A, Bharadwaj A, Saini M, Kardon G, Mathew SJ: **Myosin heavy chain- embryonic regulates skeletal muscle differentiation during mammalian development.** *Development* 2020, **147**:1-14.
143. Zieba J, Zhang W, Chong JX, Forlenza KN, Martin JH, Heard K, Grange DK, Butler MG, Kleefstra T, Lachman RS et al.: **A postnatal role for embryonic myosin revealed by MYH3 mutations that alter TGF $\beta$  signaling and cause autosomal dominant spondylometatarsal synostosis.** *Sci Rep* 2017, **7**:41803.
144. Vaughan SK, Kemp Z, Hatzipetros T, Vieira F, Valdez G: **Degeneration of proprioceptive sensory nerve endings in mice harboring amyotrophic lateral sclerosis-causing mutations.** *J Comp Neurol* 2015, **523**:2477-2494.
145. Conte A, Khan N, Defazio G, Rothwell JC, Berardelli A: **Pathophysiology of somatosensory abnormalities in Parkinson disease.** *Nat Rev Neurol* 2013, **9**:687-697.
146. Kröger S: **Proprioception 2.0: novel functions for muscle spindles.** *Curr Opin Neurol* 2018, **31**:592-598.
147. Hoffman EP, Brown RH, Kunkel LM: **Dystrophin: the protein product of the duchenne muscular dystrophy locus.** *Cell* 1987, **51**:919-928.
148. Archer JE, Gardner AC, Roper HP, Chikermane AA, Tatman AJ: **Duchenne muscular dystrophy: the management of scoliosis.** *J Spine Surg* 2016, **2**:185-194.
149. Gerwin L, Rossmannith S, Haupt C, Schultheiß J, Brinkmeier H, Bittner RE, Kröger S: **Impaired muscle spindle function in murine models of muscular dystrophy.** *J Physiol* 2020, **598**:1591-1609.
150. Barker D: **The morphology of muscle receptors.** In *Muscle Receptors*. Edited by Hunt CC. Berlin Heidelberg: Springer; 1974:1-19.
151. Shantha TR, Golarz MN, Bourne GH: **Histological and histochemical observations on the capsule of the muscle spindle in normal and denervated muscle.** *Cells Tissues Organs* 1968, **69**:632-646.
152. Ovalle WK, Dow PR: **Morphological aspects of the muscle spindle capsule and its functional significance.** In *The Muscle Spindle*. Edited by Boyd IA, Gladden MH. UK: Palgrave Macmillan; 1985:23-28.
153. Adal MN, Cheng SBC: **Capsules of duck muscle spindles.** *Cell Tissue Res* 1980, **211**:465-474.
154. Maier A, Mayne R: **Distribution of connective tissue proteins in chick muscle spindles as revealed by monoclonal antibodies: a unique distribution of brachionectin/tenascin.** *Am J Anat* 1987, **180**:226-236.
155. Lampe AK, Bushby KMD: **Collagen VI related muscle disorders.** *Med Genet* 2005, **42**:673-685.
156. Bönnemann CG: **The collagen VI-related myopathies. Ullrich congenital muscular dystrophy and Bethlem myopathy.** *Handb Clin Neurol* 2011, **101**:81-96.
157. Bonaldo P, Braghetta P, Zanetti M, Piccolo S, Volpin D, Bressan GM: **Collagen VI deficiency induces early onset myopathy in the mouse: an animal model for Bethlem myopathy.** *Hum Mol Genet* 1998, **7**:2135-2140.
158. Chen P, Cescon M, Megighian A, Bonaldo P: **Collagen VI regulates peripheral nerve myelination and function.** *FASEB J* 2014, **28**:1145-1156.